#### List of contributions

194 The roles of collagen genes in skeletal development and morphogenesis

B. R. Olsen

195 Studies of bone matrix molecules give us insights into bone remodelling

D. Heinegård

196 Gaucher disease, a paradigm for single gene defects E. Beutler

197 The chloride effect in human hemoglobin: A new kind of allosteric mechanism

M. F. Perutz

198 Scapharca dimeric hemoglobin: A new mechanism of information transfer between globin chains

E. Chiancone

199 The dimeric and co-operative myoglobin of *Nassa mutabilis*. A peculiar case

G. Geraci

200 Proteins - Paradigms of complex systems

H. Frauenfelder

204 The renaissance of myoglobin: dynamics, structure and oxygen binding control

M. Brunori

205 On and beyond O<sub>2</sub> binding: hemoglobin and myoglobin revisited

P. Saltman

206 About hemoglobins, G6PD and parasites in red cells

L. Luzzatto

# The roles of collagen genes in skeletal development and morphogenesis

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## Abstract

Molecular genetic analyses of osteochondrodysplasias in mice and humans have recently led to the identification of mutations in genes encoding structural proteins¹ growth factor receptors²,³ and sulphate transporters⁴. Further analyses of such inherited disorders, using rapid techniques for gene mapping, positional cloning and mutation detection, will undoubtedly uncover other genes that are important for skeletal development. Together with studies of transgenic mice, in which specific genes that are expressed in the skeleton are mutated, these analyses will provide insight into genes that are essential for skeletal morphogenesis.

A good example of the usefulness of the molecular genetics approach for studies of skeletal morphogenesis, is the identification of the mutation causing autosomal recessive chondrodysplasia (cho) in mice<sup>5</sup>. Homozygous cho/cho mice have severe defects in the cartilage of limbs, ribs, mandible and trachea<sup>6</sup>. They are born with cleft palate, shortened snouts, protruding tongues, short limbs, and die at birth, probably because of asphyxia as a result of tracheal collapse and/or lung hypoplasia<sup>7</sup>. Previous studies have shown that epiphyseal growth plates in cho/cho mutants do not show the normal columnar arrangement of chondrocytes as they go through maturation towards hypertrophy and contain fewer hypertrophic chondrocytes than normal growth plate cartilage<sup>8</sup>. Finally, cho/cho cartilage is mechani-

cally abnormal in that it is completely without cohesive strength, and the proteoglycan aggregates are unusually extractable. Thus, the *cho* mutation results in abnormalities both in cellular differentiation, cellular organisation and the mechanical properties of the extracellular matrix.

Since several attempts during the past 20 years to identify that *cho* gene were unsuccessful, we decided to determine the chromosomal locus of the mutation by linkage analysis. The results of such analysis showed the *cho* locus to be on mouse chromosome 3, close to the Amy1, Amy2 locus<sup>5</sup>. The gene encoding the α1(XI) chain of the quantitatively minor fibrillar collagen XI was localised to the same region, thus making *Col11a1* a candidate gene for *cho*.

To test the possibility that the *cho* abnormality could be the result of a mutation in *Coll1a1*, polyclonal antibodies against a peptide epitope within  $\alpha 1(XI)$  collagen were used to examine extracts of wild-type and *cho-cho* cartilage by Western-blotting and for immuno-histochemistry. Wild-type extracts showed, as expected, the presence of  $\alpha 1(XI)$  collagen while no reaction was seen with *cho|cho* extracts, and wild-type cartilage was positive with the anti- $\alpha 1(XI)$  antibodies while *cho-cho* cartilage was not. We concluded therefore that the *cho* mutation leads to loss of  $\alpha 1(XI)$  collagen in cartilage. Amplification of  $\alpha 1(XI)$  cDNA by RT-PCR from normal and *cho|cho* RNA, followed by nucleotide sequenc-

ing, demonstrated that the *cho* mutation is a single nucleotide deletion located about 570 nucleotides downstream of the 5' end of  $\alpha 1(XI)$  collagen mRNA. The deletion causes a shift in reading frame and results in the synthesis of a truncated  $\alpha 1(XI)$  polypeptide consisting only of the amino-terminal 1/10 of the wild-type protein. Since the levels of *cho*  $\alpha 1(XI)$  transcripts are much lower than wild-type transcripts, the levels at which such truncated peptides are synthesised must be low. This suggests that the *cho* phenotype is a loss-of-function phenotype.

What are the consequences of the loss of  $\alpha 1(XI)$  collagen in cho? Type XI collagen molecules are heterotrimers of  $\alpha 1(XI)$ ,  $\alpha 2(XI)$ , and  $\alpha 3(XI)$  collagen chains and the absence of  $\alpha 1(XI)$  must lead to a deficiency of type XI collagen molecules. Collagen fibrils in cho/cho cartilage are abnormally thick, thus suggesting that type XI collagen molecules are essential for the formation of a network of thin collagen fibrils in cartilage. This could explain the abnormal extractability of proteoglycans form cho/cho cartilage and the lack of cohesiveness, in that fewer, albeit thicker fibrils would provide less of a physical entrapment of large proteoglycan aggregates. Alternatively, type XI collagen molecules or sub-domains may bind to proteoglycans in normal cartilage, and the loss of such binding sites in cho/cho cartilage could result in decreased tissue cohesiveness.

What the reasons may be for the defect in chondrocyte differentiation seen in *cho/cho* growth plate regions is not clear. Perhaps type XI collagen molecules along collagen fibrils contain binding sites for chondrocyte matrix receptors that play a role in signalling pathways controlling chondrocyte differentiation. Alternatively, type XI molecules may directly (or indirectly) bind growth factors that are essential for inducing chondrocyte differentiation. Whatever the reasons may be, however, it is clear that the *cho* mice provide a system for studying how extracellular matrix regulates cell differentiation, cellular organisation and tissue cohesiveness. In addition, the mice provide an animal model for studying molecular mechanisms in human disorders due to

mutations in type XI collagen chains. The first examples of such disorders are osteochondrodysplasias in two Dutch kindreds linked to the COL11A2 locus. In one family with an autosomal dominant form of the Stickler syndrome, affected individuals have mild spondyloepiphyseal dysplasia, osteoarthritis, and sensorineural hearing loss<sup>9</sup>. In this family a splice donor site mutation results in 'in-frame' exon skipping within  $\alpha 2(XI)$  collagen transcripts<sup>10</sup>. In a second family with similar, but more severe characteristics inherited as an autosomal recessive abnormality we have also shown linkage to COL11A2<sup>10</sup>. These results suggest that mutations in collagen XI genes are associated with a spectrum of abnormalities in human skeletal development and support the conclusion based on the *cho* studies, that collagen XI is essential for skeletal morphogenesis.

- 1 Warman, M. L., Abbott, M., Apte, S. S., Hefferon, T., McIntosh, I., Cohn, D., Hecht, J., Olsen, B. R., and Francomano, C.A., Nature Genet. 5 (1993) 79.
- 2 Shiang, R., Thompson, L. M., Zhu, Y.-Z., Church, D. M., Fiedler, T. J., Bocian, M., Winokur, S. T., and Wasmuth, J. J., Cell 78 (1994) 335.
- 3 Reardon, W., Winter, R. M., Rutland, P., Pulleyn, L. J., Jones, B. M., and Malcolm, S., Nature Genet. 8 (1994) 98.
- 4 Hastbacka, J., de la Chapelle, A., Mahtani, M. M., Clines, G., Reeve-Daly, M. P., Daly, M., Hamilton, B. A., Kusumi, K., Trivedi, B., Weaver, A., Coloma, A., Lovett, M., Buckler, A., Kaitila, I., and Lander, E. S., Cell *78* (1994) 1073.
- 5 Li, Y., Lacerda, D. A., Warman, M. L., Beier, D. R., Yoshioka, H., Ninomiya, Y., Oxford, J. T., Morris, N. P., Andrikopoulos, K., Ramirez, F., Wardell, B. B., Lifferth, G. D., Teuscher, C., Woodward, S. R., Taylor, B. A., Seegmiller, R. E., Olsen, B. R., Cell February 1995.
- 6 Seegmiller, R. E., Fraser, F. C., and Sheldon, H., J. Cell Biol. 48 (1971) 580.
- 7 Seegmiller, R. E., Ferguson, C. C., and Sheldon, H., J. Ultrastr. Res. 38 (1972) 288.
- 8 Monson, C. B., and Seegmiller, R. E., J. Bone Jt Surg. *63*A (1981) 637.
- 9 Brunner, H. G., van Beersum, S. E. C., Warman, M. L., Olsen, B. R., Ropers, H.-H., and Mariman, E. C. M., Hum. molec. Genet. 3 (1994) 1561.
- 10 Vikkula, M., Mariman, E. C. M., Lui, V. C. H., Zhidkova, N. I., Tiller, G. E., Goldring, M. B., van Beersum, S. E. C., Malefijt, M., van den Hoogen, F. H. J., Ropers, H.-H., Mayne, R., Cheah, K. S. E., Olsen, B. R., Warman, M. L., and Brunner, H. G., Cell February 1995.

## Studies of bone matrix molecules give us insights into bone remodelling

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#### Abstract

Bone is one of the most dynamic tissues, with constant remodelling and repair of minor defects during the whole life cycle. These processes have key roles in replacing fatigued, non-functional tissue with fresh tissue which has the proper mechanical properties. In this process, a balance between breakdown of less functional tissue and new tissue production is a prerequisite for tissue integrity and function. One of the major health problems in the elderly, i.e. osteoporosis and